

Towards a global understanding of protein production in prokaryotes

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PhD Defense

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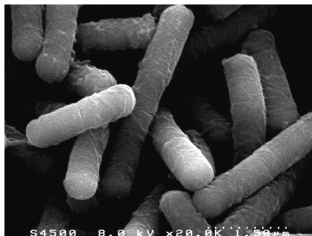


Outline:

- 1 Introduction
- 2 Stochastic model of Gene Expression: Single-Protein
- 3 Stochastic model of Gene Expression: Multi-Protein

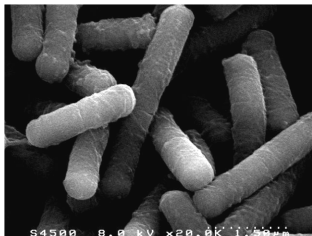
Central role of proteins in prokaryotes

- Proteins are the core of biologic processes: *enzymes*, DNA replication machinery, ...
- ~ 50% of the bacteria dry weight
- ~ 3.5 millions of proteins in each cell
- ~ 2000 types of proteins produced at any time at any growth condition
- proteins ranging from few dozens up to 10^5



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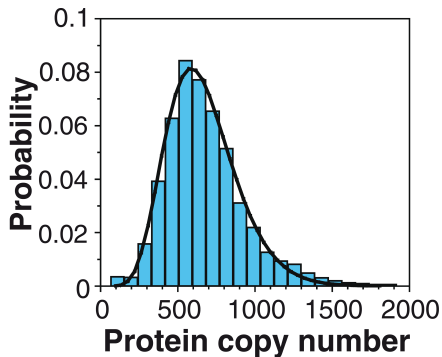
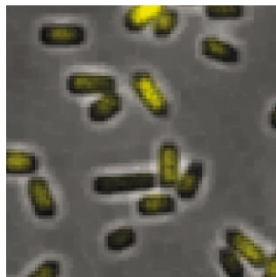


A highly consuming process:

- at each generation, the bacterium has to duplicate all proteins
- more than 85% of cell resources

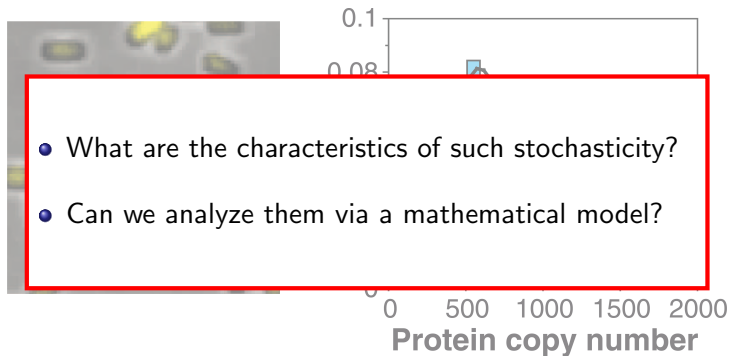
Stochasticity in protein production: experimental viewpoint

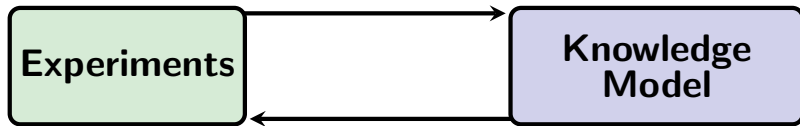
E. Coli – ADk cytoplasm protein¹

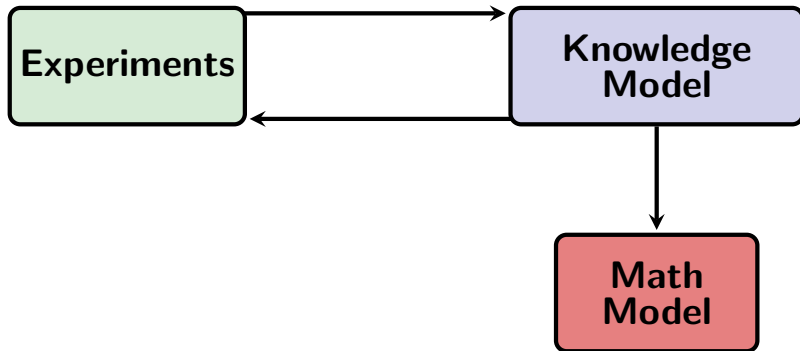


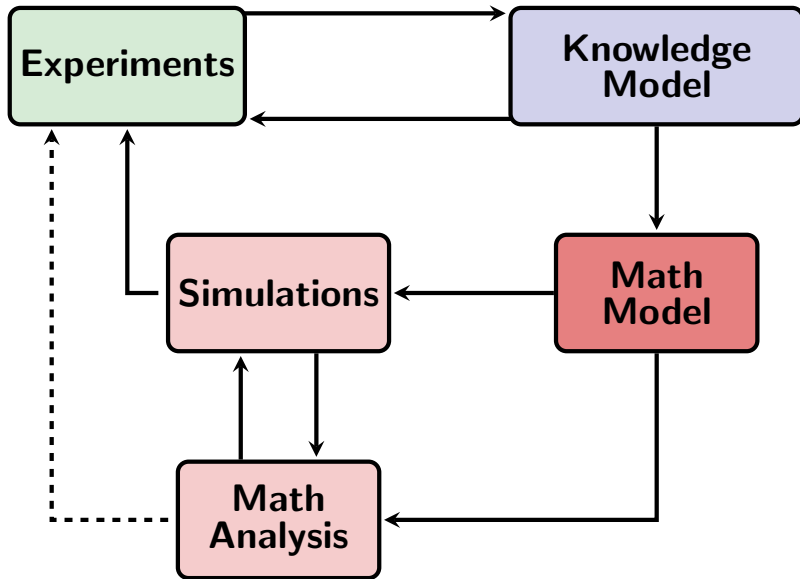
¹Yuichi Taniguchi et al. *Science* (2010), pp. 533–538.

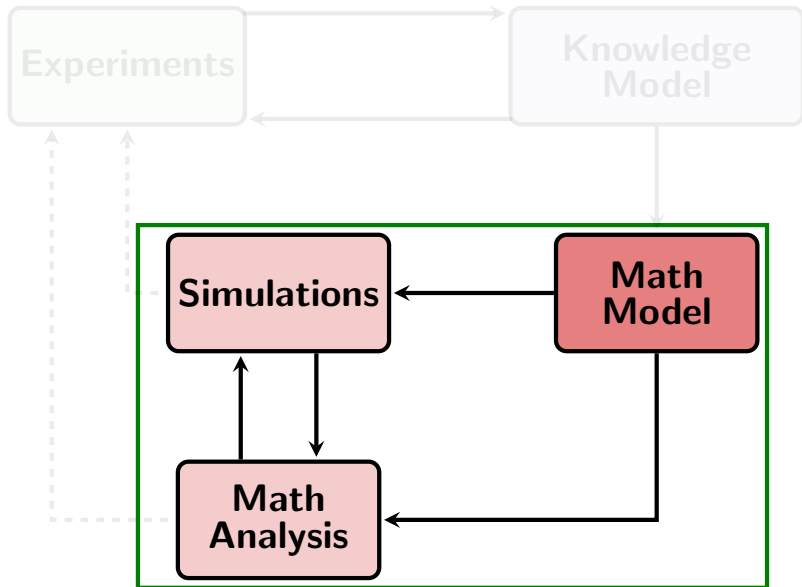
Stochasticity in protein production: experimental viewpoint











Part I

Stochastic model of Gene Expression: Single-Protein

4-Stage model: activation



Active Gene

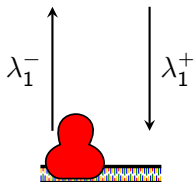


Inactive Gene

$$Y(t) \in \{0, 1\}$$

Gene status

4-Stage model: activation



$$Y(t) \in \{0, 1\}$$

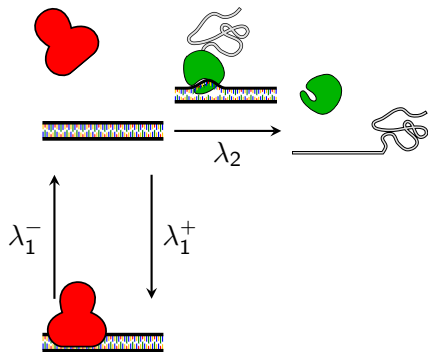
Gene status

Exponential assumption

- diffusion in a stiff medium
- thermal noise
- biochemical reactions (in general): encounter of macromolecules

exponentially distributed duration

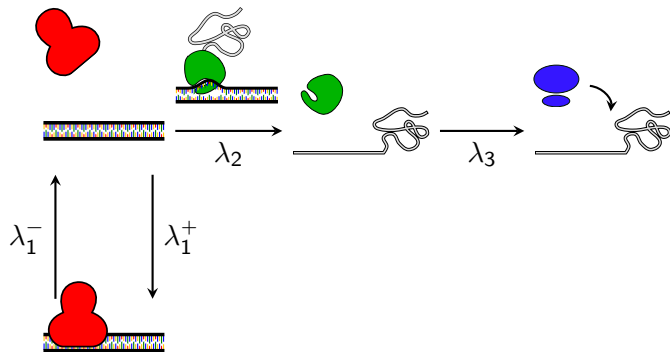
4-Stage model: transcription



$$Y(t) \in \{0, 1\} \longrightarrow M(t) \in \mathbb{N}$$

Messenger

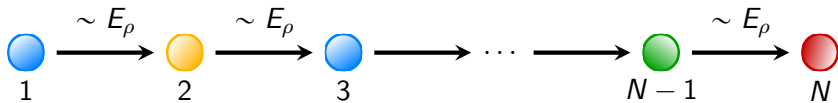
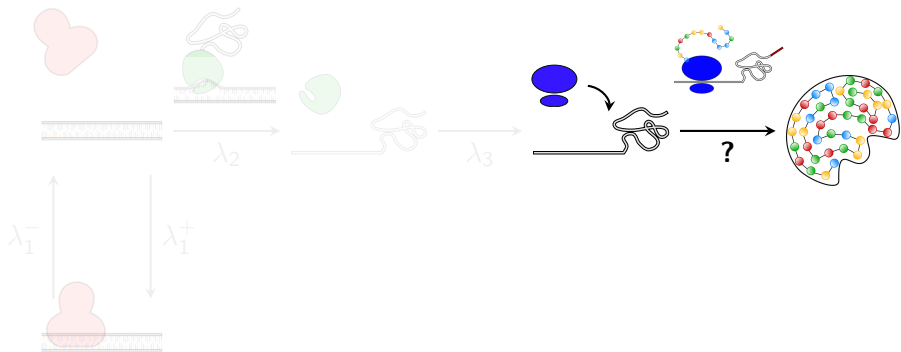
4-Stage model: translation initiation



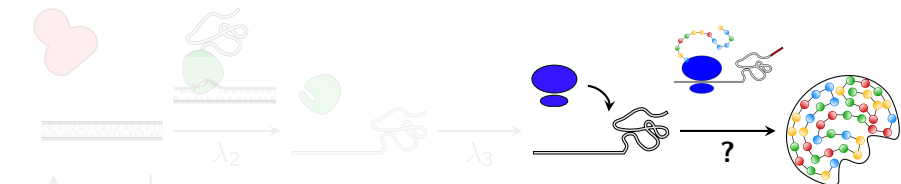
$$Y(t) \in \{0, 1\} \longrightarrow M(t) \in \mathbb{N} \longrightarrow R(t) \in \mathbb{N}$$

Ribosome

4-Stage model: translation completion

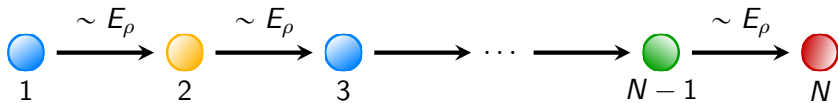


4-Stage model: translation completion

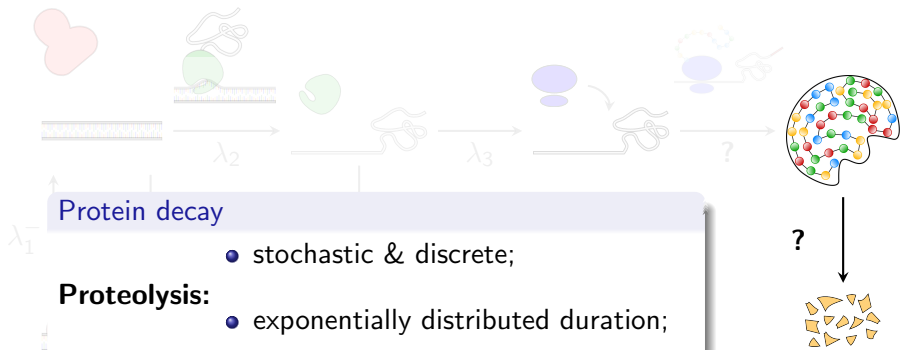


Protein Elongation

- exponentially distributed elementary steps
- **Erlang distributed** elongation $T^{(el.)} \sim \text{Erl}(N, \rho)$
($N \approx 400$ in average)



4-Stage model



Protein decay

- stochastic & discrete;

Proteolysis:

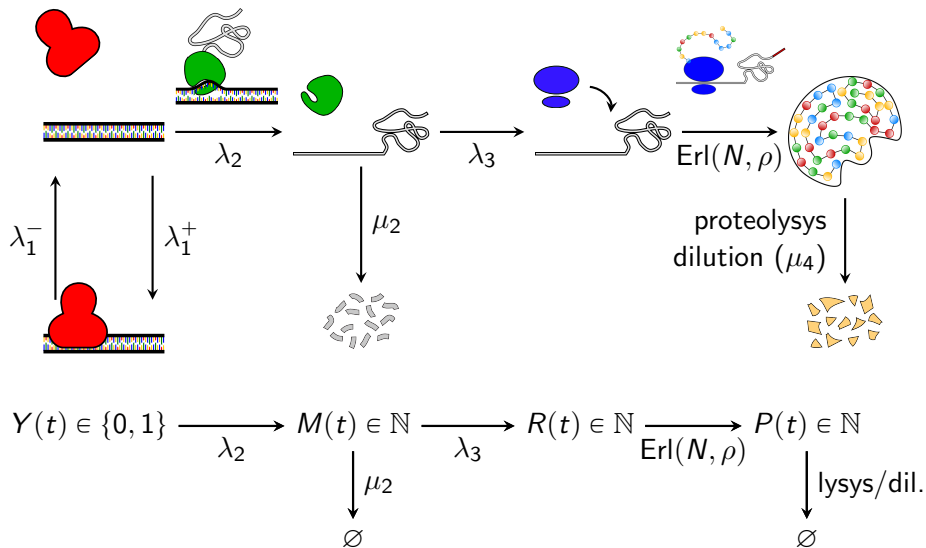
- exponentially distributed duration;

-
- deterministic & continuous;

Dilution:

- main decay mechanism;

4-Stage model

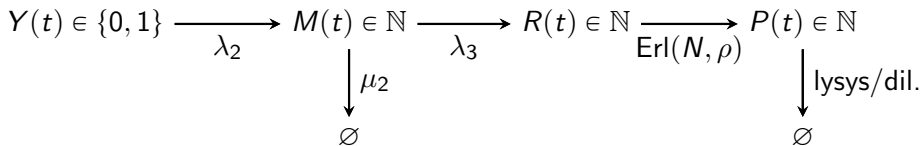


4-Stage model

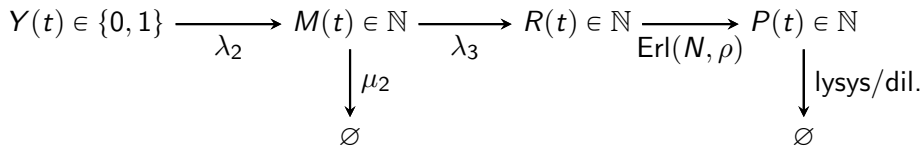
Protein production system:

- fixed number of proteins required
- more than 85% of resources

→ variance is a **key parameter**



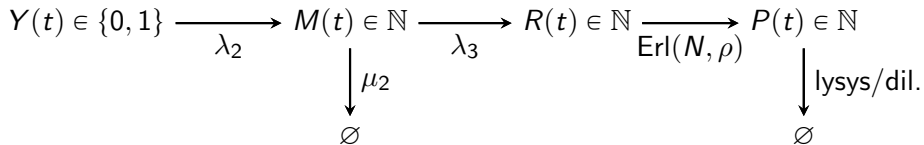
4-Stage model



Goals:

- ▶ characterize **mean** and **variance** of proteins P at equilibrium
- ▶ identify the role of the different parameters

4-Stage model



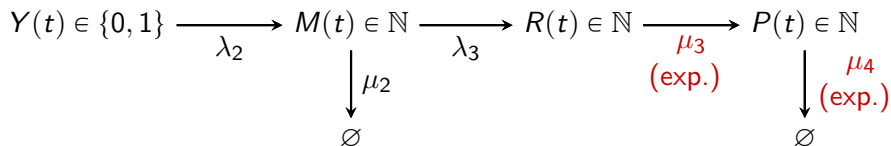
Goals:

- ▶ characterize **mean** and **variance** of proteins P at equilibrium
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Why is it difficult to achieve these goals?

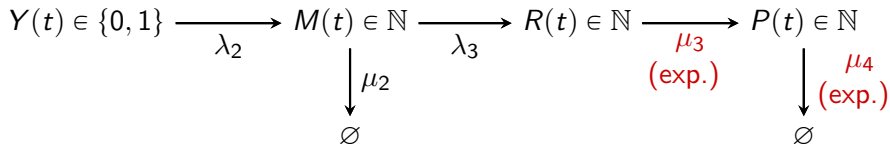
The need for a new framework to overcome the limitations of the classic approach.

Limits of classic approach



- *Assumption*: each step has **exponentially distributed** duration
- *Markovian* description of protein production

Limits of classic approach



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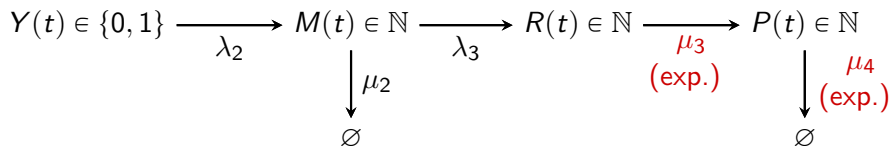
30 years of gene expression:

Berg (1978), Rigney (1979), Swain (2002), Paulsson (2005), ...

Original motivation for Math Models:

Lack of experimental data

Limits of classic approach



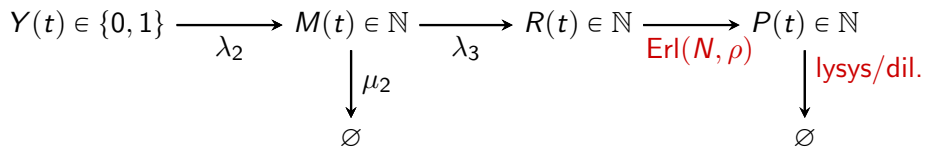
- *Assumption*: each step has **exponentially distributed** duration
- *Markovian* description of protein production

Results:

- *reference model* for biologists
- quantitative characterization of protein fluctuations

$$\text{var}(P) = \mathbb{E}[P] \left[1 + \frac{\lambda_3 \mu_3 (\mu_2 + \mu_3 + \mu_4)}{(\mu_2 + \mu_3)(\mu_2 + \mu_4)(\mu_3 + \mu_4)} \right]$$

Limits of classic approach

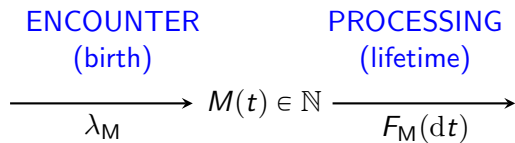


Limitations

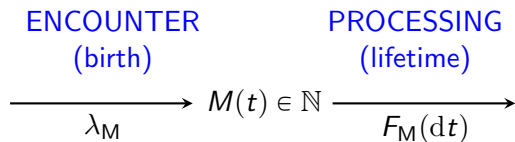
Classic framework cannot be used in non-Markovian description

How to include non exponential steps?
Marked Poisson Point Process (MPPP) framework

Explanatory model



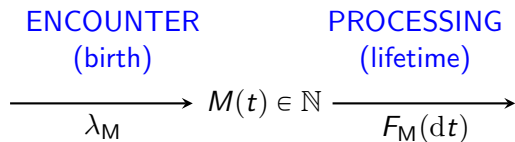
Explanatory model



Assumptions:

- births (s_n) follow a **Poisson process** of parameter λ_M
- lifetimes (σ_n) with distribution $F_M(dt)$ (**mark**)

Explanatory model



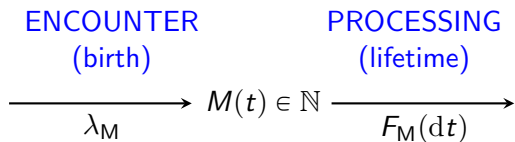
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- births (s_n) follow a **Poisson process** of parameter λ_M
- lifetimes (σ_n) with distribution $F_M(dt)$ (**mark**)

$\mathcal{M} = (s_n, \sigma_n)$

 marked Poisson point process on $\mathbb{R} \times \mathbb{R}_+$
 with intensity $\mu_{\mathcal{M}} = \lambda_M du \otimes F_M(dv)$

Explanatory model

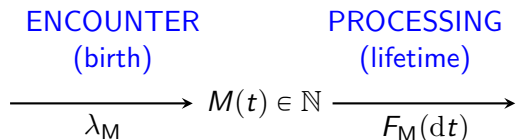


At equilibrium, $M(\infty)$ is a functional of Poisson process \mathcal{M} :

$$M(\infty) = \mathcal{M}(\mathbb{1}_{\{u \leq 0 \leq u+v\}}) = \int_{\mathbb{R} \times \mathbb{R}_+} \mathbb{1}_{\{u \leq 0 \leq u+v\}} \mathcal{M}(du, dv)$$

where $\mathcal{M}(f) = \sum_n f(s_n, \sigma_n) = \int_{\mathbb{R} \times \mathbb{R}_+} f(x, y) \mathcal{M}(dx, dy)$

Explanatory model



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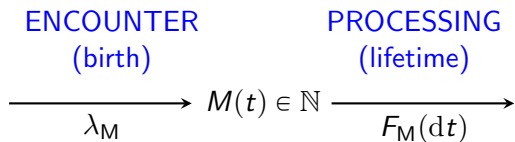
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For any nice $f : \mathbb{R} \times \mathbb{R}_+ \rightarrow \mathbb{R}_+$, the **Laplace transform** of \mathcal{M} is

$$\mathcal{L}_{\mathcal{M}}(f) = \mathbb{E} \left[e^{-\mathcal{M}(f)} \right] = \exp \left(- \int \left(1 - e^{-f(u,v)} \right) \lambda_M du F_M(dv) \right)$$

Explanatory model



At equilibrium, $M(\infty)$ is a functional of Poisson process \mathcal{M} :

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Proposition

$$\begin{aligned}
 \mathbb{E}[M] &= \lambda_M \mathbb{E}[\sigma], & \sigma &\sim F_M(dv) \\
 \text{var}(M) &= \mathbb{E}[M]
 \end{aligned}$$

Proteins

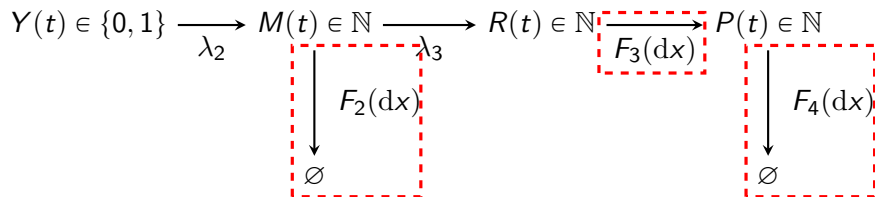
In the case of proteins

- $\mathcal{P} = \left(s_n, \sigma_n, \mathcal{N}_{\tilde{\lambda}}^{\sigma_n} \right)$ is a Poisson process with intensity measure $\mu_{\mathcal{P}}$
- the **mark** is now $\left(\sigma_n, \mathcal{N}_{\tilde{\lambda}}^{\sigma_n} \right)$
- $\mathcal{N}_{\tilde{\lambda}}^{\sigma_n}$ is a Poisson process on $\mathbb{R} \times \mathbb{R}_+ \times \mathbb{R}_+$ associated to a mRNA born in s_n

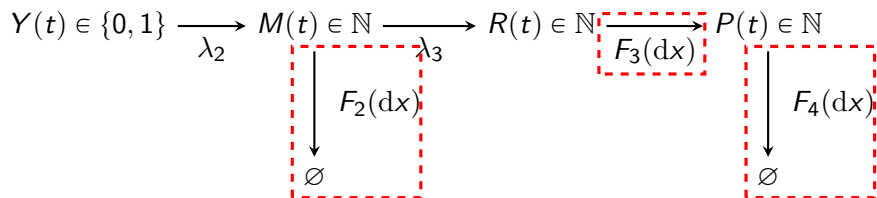
same approach but more complicated

MPPP & Gene Expression

General formulas of protein statistics



General formulas of protein statistics



Mean:

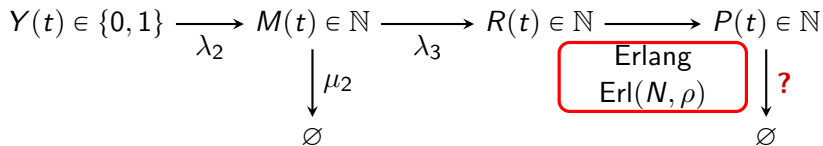
$$\mathbb{E}(P) = \lambda_2 \lambda_3 \mathbb{E}(F_2) \mathbb{E}(F_4)$$

Variance (always active gene):

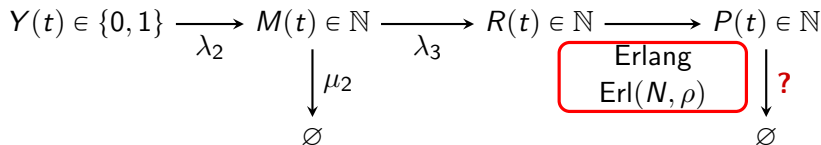
$$\text{var}(P) = \mathbb{E}(P) + \lambda_2 \lambda_3^2 \int_{\mathbb{R} \times \mathbb{R}_+} \mathbb{1}_{\{s \leq 0\}} \left[\int_{\mathbb{R}_+^2} \mathbb{1}_{\{-(u+s+t) \leq y \leq -(u+s)\}} \bar{F}_4(u) \, du F_3(dy) \right]^2 ds F_2(dt)$$

Application

Application



Protein decay: Proteolysis & Dilution



Protein decay:

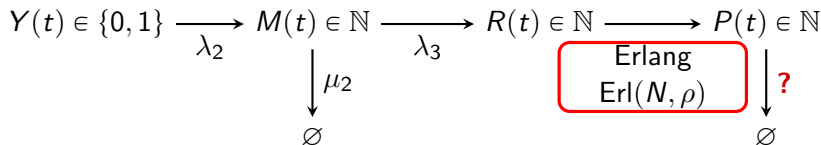
Proteolysis

- ① stochastic and discrete nature (*exponential*);
- ② used classically to describe protein decay

Dilution

-
- ① main phenomenon of decay (in general);
 - ② deterministic and continuous nature;
 - ③ understudied phenomenon in models;

Protein decay: Proteolysis & Dilution

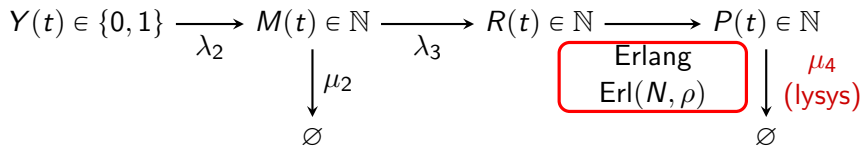


Protein decay:

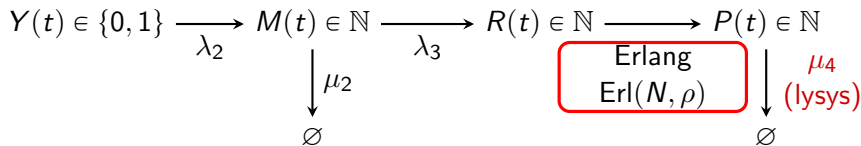
MPPP framework can describe both *proteolysis* and *dilution*

- *Proteolysis*: exponentially distributed of parameter μ_4
- *Dilution*: deterministic, with growth rate parameter ν

Application: protein elongation & proteolysis



Application: protein elongation & proteolysis

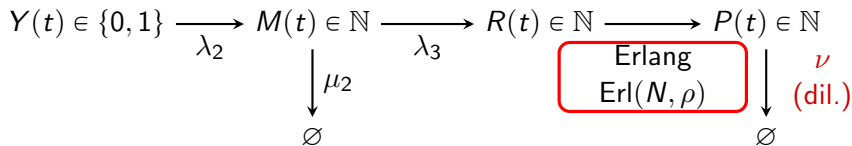


Protein variance: Erlang elongation & proteolysis

$$\begin{aligned}
 \text{var}_{\text{LYSIS}}^{(\text{Erl})}(P) = \mathbb{E}[P] & \left[1 + \frac{2\lambda_3\mu_2}{\mu_2^2 - \mu_4^2} \frac{\rho^{2N}}{(N-1)!} \right. \\
 & \times \left(\frac{\mu_2}{(\rho^2 - \mu_4^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_4^2}\right) ds \right. \\
 & \left. \left. - \frac{\mu_4}{(\rho^2 - \mu_2^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_2^2}\right) ds \right) \right]
 \end{aligned}$$

$Q(N, s)$ is the complementary cumulative distribution function of $\text{Erl}(N, 1)$, N is the number of amino acids and ρ is the rate of elongation of each amino acid

Application: protein elongation & dilution



Protein variance: Erlang elongation & dilution

$$\begin{aligned}
 \text{var}_{\text{DIL.}}^{(\text{Erl})}(P) = \mathbb{E}[P] & \left[\frac{1}{2} + \frac{2\lambda_3\mu_2}{\mu_2^2 - \nu^2} \frac{\rho^{2N}}{(N-1)!} \right. \\
 & \times \left(\frac{\mu_2}{(\rho^2 - \nu^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \nu^2}\right) ds \right. \\
 & \left. \left. - \frac{\nu}{(\rho^2 - \mu_2^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_2^2}\right) ds \right) \right]
 \end{aligned}$$

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Proteolysis decay [$\mu_4 = \nu$]

$$\begin{aligned} \text{var}_{\text{LYSIS}}^{(\text{Erl})}(P) = \mathbb{E}[P] & \left[1 + \frac{2\lambda_3\mu_2}{\mu_2^2 - \nu^2} \frac{\rho^{2N}}{(N-1)!} \right. \\ & \times \left(\frac{\mu_2}{(\rho^2 - \nu^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \nu^2}\right) ds \right. \\ & \left. \left. - \frac{\nu}{(\rho^2 - \mu_2^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_2^2}\right) ds \right) \right] \end{aligned}$$

Dilution decay

$$\begin{aligned} \text{var}_{\text{DIL.}}^{(\text{Erl})}(P) = \mathbb{E}[P] & \left[\frac{1}{2} + \frac{2\lambda_3\mu_2}{\mu_2^2 - \nu^2} \frac{\rho^{2N}}{(N-1)!} \right. \\ & \times \left(\frac{\mu_2}{(\rho^2 - \nu^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \nu^2}\right) ds \right. \\ & \left. \left. - \frac{\nu}{(\rho^2 - \mu_2^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_2^2}\right) ds \right) \right] \end{aligned}$$

Proteolysis decay [$\mu_4 = \nu$]

$$\text{var}_{\text{LYSIS}}^{(\text{Erl})}(P) = \mathbb{E}[P] \left[1 + \frac{2\lambda_3\mu_2}{\mu_2^2 - \nu^2} \frac{\rho^{2N}}{(N-1)!} \times \left(\frac{\mu_2}{(\rho^2 - \nu^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \nu^2}\right) ds \right. \right. \\ \left. \left. - \frac{\nu}{(\rho^2 - \mu_2^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_2^2}\right) ds \right) \right]$$

True for any distribution

This result is general since it holds true for any choice of distributions

Dilution decay

$$\text{var}_{\text{DIL.}}^{(\text{Erl})}(P) = \mathbb{E}[P] \left[\frac{1}{2} + \frac{2\lambda_3\mu_2}{\mu_2^2 - \nu^2} \frac{\rho^{2N}}{(N-1)!} \times \left(\frac{\mu_2}{(\rho^2 - \nu^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \nu^2}\right) ds \right. \right. \\ \left. \left. - \frac{\nu}{(\rho^2 - \mu_2^2)^N} \int_{\mathbb{R}_+} s^{N-1} e^{-s} Q\left(N, \frac{s}{\rho^2 - \mu_2^2}\right) ds \right) \right]$$

Summary

MPPP Approach:

- more appropriate description of gene expression
- mix of different probability distributions
- mix of deterministic/stochastic processes
- analytic formulas of mean and variance
- impact of dilution/proteolysis

... but

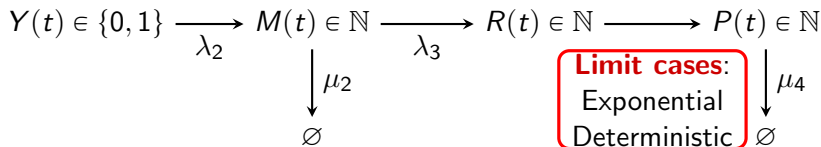
Despite the obtained formulas can be still studied numerically, we have lost a bit of intuition of the formulas derived in the classic approach, i.e.

$$\text{var}(P) = \mathbb{E}[P] \left[1 + \frac{\lambda_3 \mu_3 (\mu_2 + \mu_3 + \mu_4)}{(\mu_2 + \mu_3)(\mu_2 + \mu_4)(\mu_3 + \mu_4)} \right]$$

Idea:

study two specific cases leading to variance formulas “explicit” with respect to model parameters

Protein elongation: “limit cases”



“Limit cases”:

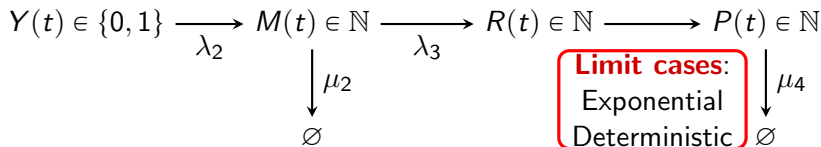
Exponential:

- classic assumption
- high fluctuations on elongation step

Deterministic:

- no fluctuations of elongation duration
- closer to reality (on average ≈ 400 a.a.)

Protein elongation: “limit cases”



Limit cases

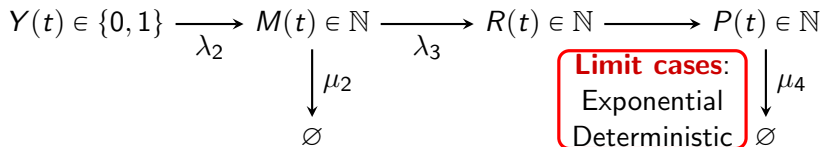
Exponential elongation:

$$\text{var}_{\mathbb{E}}(P) = \mathbb{E}(P) \left[1 + \frac{\lambda_3 \mu_3 (\mu_2 + \mu_3 + \mu_4)}{(\mu_2 + \mu_3)(\mu_2 + \mu_4)(\mu_3 + \mu_4)} + \frac{\lambda_2 \lambda_3 (1 - \delta_+) \mu_3 \mu_4^2}{(\Lambda + \mu_2)(\mu_4^2 - \mu_3^2)} \times \left(\frac{\Lambda + \mu_2 + \mu_3}{\mu_3 (\mu_2 + \mu_3) (\Lambda + \mu_3)} - \frac{\Lambda + \mu_2 + \mu_4}{\mu_4 (\mu_2 + \mu_4) (\Lambda + \mu_4)} \right) \right],$$

Deterministic elongation:

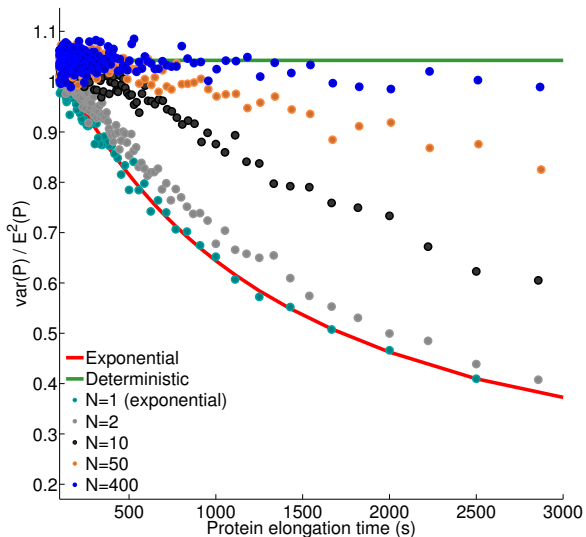
$$\text{var}_{\mathbb{D}}(P) = \mathbb{E}(P) \left[1 + \frac{\lambda_3}{\mu_2 + \mu_4} + \frac{\lambda_2 \lambda_3 (1 - \delta_+) (\Lambda + \mu_2 + \mu_4)}{(\mu_2 + \mu_4) (\Lambda + \mu_2) (\Lambda + \mu_4)} \right]$$

Protein elongation: “limit cases”



Counter-intuitive theoretical result: $\text{var}_D(P) \geq \text{var}_E(P)$

Erlang elongation: simulations

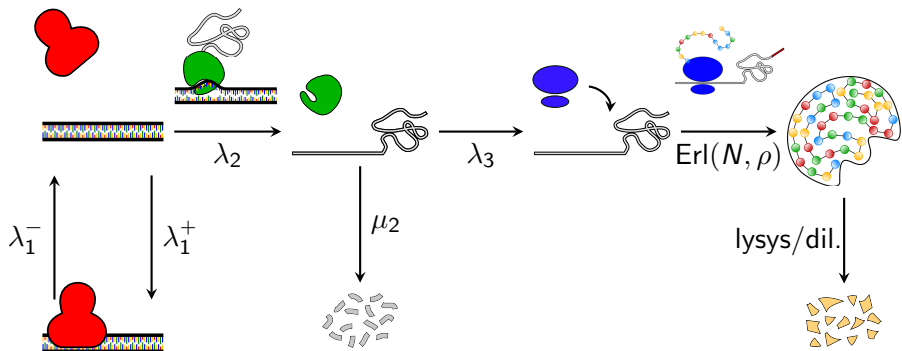


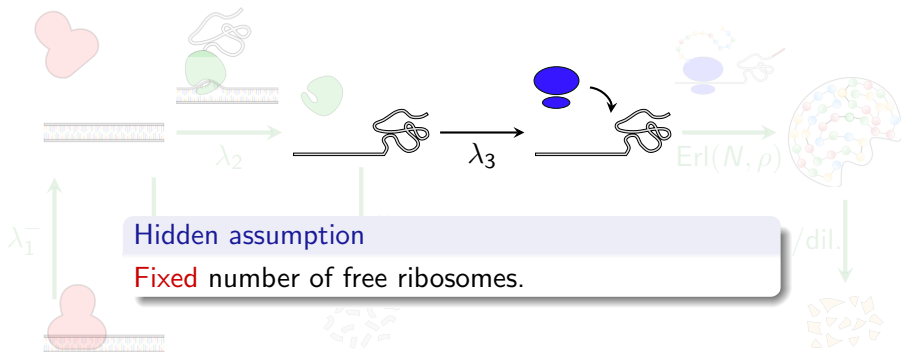
Conclusions

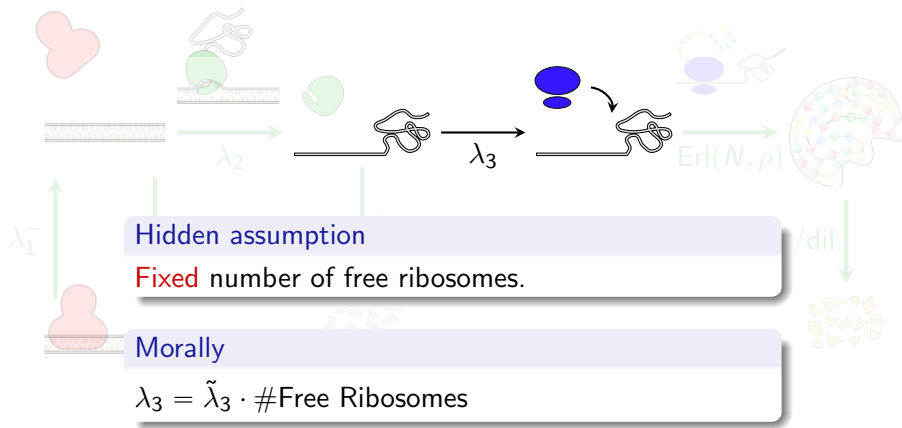
- analysis and proof of the correct assumptions of gene expression
- analytic formulas for any distribution
- two “explicit” formulas (estimation of impact of different choices)
- counter-intuitive: $\text{var}_{\text{DET}}(P) \geq \text{var}_{\text{EXP}}(P)$
- classic models underestimate protein variance
- (MPPP) appropriate math tool to describe cell stochastic processes
(**Encounter + Processing**)

Part II

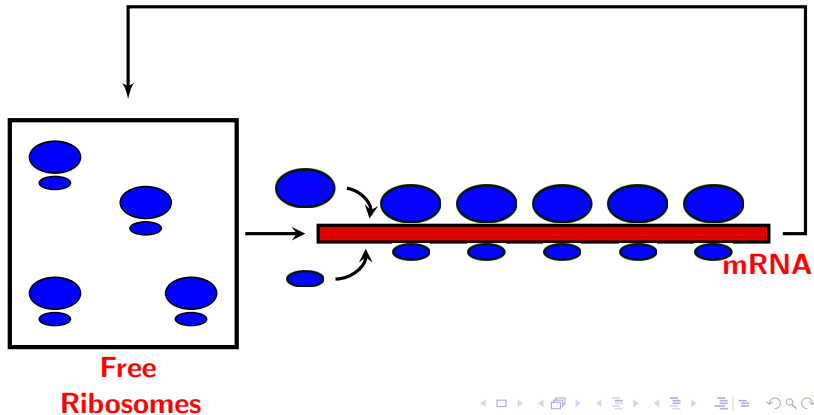
Stochastic model of Gene Expression: Multi-Protein







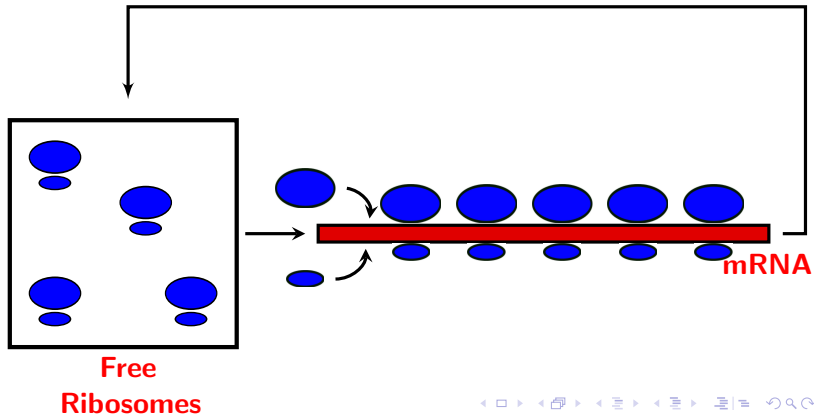
Free ribosomes



Free ribosomes

Free ribosomes

- few *free ribosomes* available
- random variable



Free ribosomes

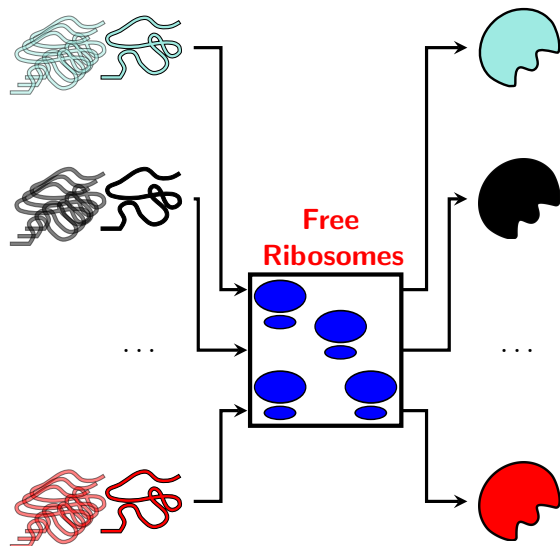


...

...



Free ribosomes



Ribosomes

- expensive
- limited number (N)

Numbers

- $N \approx 40.000$ total ribosomes
- ≥ 100.000 ribosomes required

Objective:

Characterize the law of *free ribosomes* and the state of the system

A simple model:

- P values of concentrations of proteins
- N total (fixed) number of ribosomes
- K_p (fixed) number of mRNAs of class p
- C_p capacity of mRNAs of class p

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Quantities of interest

- $X_{p,k}^N(t)$ number of ribosomes attached on the k^{th} messenger of class p
(**active ribosomes**)

- $R(t) = N - \sum_{p,k} X_{p,k}(t)$ **free ribosomes**

for $p = 1, \dots, P$, $k = 1, \dots, K_p$

Reversible Markov process:

$$(\mathcal{X}(t)) = \{(X_{p,k}(t)) : 1 \leq p \leq P, 1 \leq k \leq K_p\}$$

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Proposition (Invariant distribution)

The invariant distribution π of $(\mathcal{X}(t))$ is given by

$$\pi(x) = \frac{1}{Z} \frac{1}{R(x)!} \prod_{p=1}^P \prod_{k=1}^{K_p} \rho_p^{x_{p,k}} \quad x \in \mathcal{S}$$

where $\rho_p = \lambda_p / \mu_p$, $R(x)$ free ribosomes.

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- ▶ product form invariant distribution
- ▶ complicated normalization constant

Large scale model

Scaling

- N ribosomes (N large)
- $K_p^N \approx N\beta_p$, for $p = 1, \dots, P$

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Without constraints on ribosomes let G_p number of ribosomes attached at equilibrium on a mRNA of class (p, k)

Saturation Condition:

$$\sum_{p=1}^P K_p^N \mathbb{E}(G_p) > N$$

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Large scale model

Ingredients

Limiting regime: $K_p^N \approx N\beta_p$

Saturation condition:
$$\sum_{p=1}^P \beta_p \rho_p \frac{C_p \rho_p^{C_p+1} - (C_p + 1) \rho_p^{C_p} + 1}{(1 - \rho_p)(1 - \rho_p^{C_p+1})} > 1$$

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Theorem (Free ribosomes limiting regime)

At equilibrium, as $N \rightarrow \infty$, the nb. of free ribosomes is *Poisson* with parameter $\gamma(\underline{C})$, solution of equation

$$\sum_{p=1}^P \beta_p \rho_p \gamma \frac{C_p \rho_p^{C_p+1} \gamma^{C_p+1} - (C_p + 1) \rho_p^{C_p} \gamma^{C_p} + 1}{(1 - \rho_p \gamma)(1 - \rho_p^{C_p+1} \gamma^{C_p+1})} = 1,$$

$\underline{C} = (C_p)$ and $\rho_p = \lambda_p / \mu_p$.

Large scale model

Ingredients

Limiting regime: $K_p^N \approx N\beta_p$

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$$\sum_{p=1}^P \beta_p \rho_p \frac{C_p \rho_p^{C_p+1} - (C_p + 1) \rho_p^{C_p} + 1}{(1 - \rho_p)(1 - \rho_p^{C_p+1})} > 1$$

Theorem (Active ribosomes limiting regime)

At equilibrium, as $N \rightarrow \infty$,

- the r.v. $(X_{p,k}^N)$ are independent
- the system $(X_{p,k}^N)$ behaves as if there were no constraints:
 λ_p is now replaced by $\lambda_p \gamma$

Large scale model

Theorem (Free ribosomes limiting regime)

At equilibrium, as $N \rightarrow \infty$, the nb. of free ribosomes is Poisson with parameter $\gamma(\underline{C})$, solution y of equation

$$\sum_{p=1}^P \beta_p \rho_p \gamma \frac{C_p \rho_p^{C_p+1} \gamma^{C_p+1} - (C_p + 1) \rho_p^{C_p} \gamma^{C_p} + 1}{(1 - \rho_p \gamma)(1 - \rho_p^{C_p+1} \gamma^{C_p+1})} = 1.$$

Proof.

- Mean-Field Limit Setting
- Change of Probability Measure
- Local Central Limit Theorem



Conclusions

Multi-protein model:

- *competition for common resources (free ribosomes)*
- Markov-model with limited number of ribosomes and limited mRNA capacity

Results:

- *free ribosomes* are Poisson distributed (asymptotic regime)
- *active ribosomes* are independent in the asymptotic regime
- interactions described by fixed-point equation

Perspectives

- model with varying numbers of messengers
- *polymerases*: similar approach, but... some peculiarity:
 - several types of polymerases (different *gamma factors*)
 - stand-by polymerases on DNA (*buffering?*)
 - polymerase diffusion transport mechanism (not only 3D diffusion?)
- extrinsic noise: better understanding of the role of ribosomes/polymerases on the protein production

Thanks.